

**ROLE OF SULFONAMIDE MOIETY IN NON-PROSTANOID TXA<sub>2</sub> RECEPTOR  
ANTAGONIST KT2-962 : MODIFICATIONS OF THIS MOIETY  
AND THE RESULTING ACTIVITIES**

Masayuki Yokota,\* Kenji Imamaki, Satoko Uchibori, Masahiro Kondo, Kazuhiro Kosakai  
and Tsuyoshi Tomiyama

Kotobuki Research Laboratories, Kotobuki Seiyaku Company, Ltd., 6351 Sakaki-machi,  
Nagano-ken, 389-06, Japan

*(Received in USA 19 February 1993; accepted 15 April 1993)*

**Abstract:** Modification of sulfonamide moiety in non-prostanoid thromboxane A<sub>2</sub> (TXA<sub>2</sub>) receptor antagonist, KT2-962 with double amide, sulfonamide-amide, (thio)semicarbazone, inverse sulfonamide and N-sulfonylcarboxamide is described. Unlike prostanoid TXA<sub>2</sub> antagonists, the importance of sulfonamide moiety for the activity of non-prostanoid TXA<sub>2</sub> receptor antagonist, KT2-962 was confirmed.

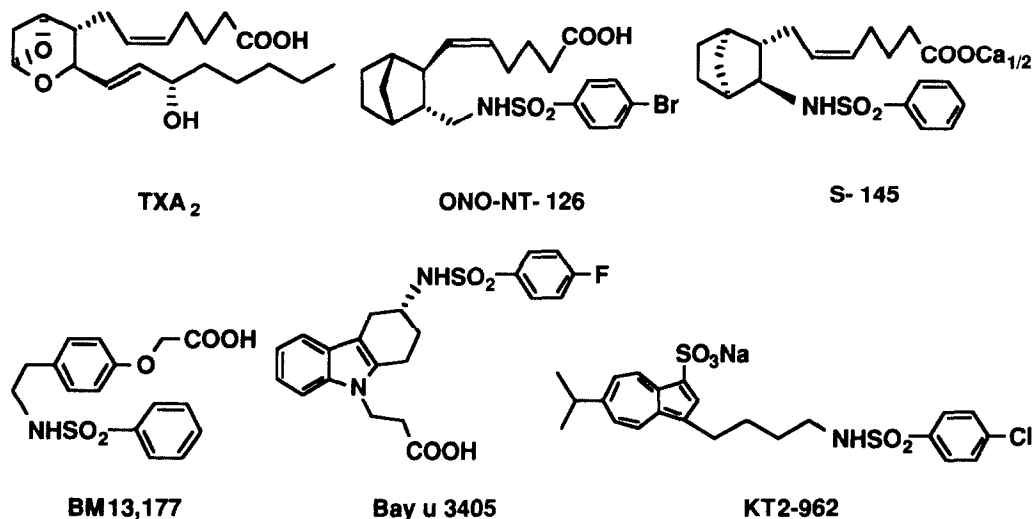
TXA<sub>2</sub> is known to contract different types of smooth muscles and to activate circulating blood cells. TXA<sub>2</sub> usually constricts blood vessels, contracts respiratory smooth muscles and induces aggregation or adhesion of platelets, erythrocytes and perhaps endothelial cells.<sup>1</sup> Agents which modulate the actions of TXA<sub>2</sub> are classified as TXA<sub>2</sub> synthetase inhibitors and TXA<sub>2</sub> receptor antagonists.

In general, TXA<sub>2</sub> synthetase inhibitors have been disappointing in terms of their efficacy in circulatory disorders in humans. However, TXA<sub>2</sub> receptor antagonists may be more useful than TXA<sub>2</sub> synthetase inhibitors, because these compounds also antagonize the effect of endoperoxides and they do not lead to accumulation of endoperoxide intermediate. Therefore, TXA<sub>2</sub> receptor antagonists appear to have greater potential as therapeutics.<sup>2</sup>

TXA<sub>2</sub> receptor antagonists are classified as prostanoid TXA<sub>2</sub> receptor antagonists (P-TRAs) and non-prostanoid TXA<sub>2</sub> receptor antagonists (NP-TRAs). P-TRAs are reported to have partial agonistic activities<sup>3</sup> and NP-TRAs are considered not to have partial agonistic activities.<sup>4</sup>

In the search for P-TRAs and NP-TRAs, a number of compounds (ONO-NT-126,<sup>5</sup> S-145,<sup>6</sup> BM13,177,<sup>7</sup> Bay u 3405<sup>8</sup>), having sulfonamide moiety, have been reported and sulfonamide seems to be one of the active functionalities in P-TRAs and NP-TRAs. Several attempts to modify allylic alcohol functionality in P-TRAs were made and replacements of the allylic alcohol moiety with double amide<sup>9</sup> and semicarbazone<sup>10</sup> on omega chain have been described.

We have previously reported the synthesis of a series of azulene derivatives, one of which is 6-isopropyl-3-[4-(p-chlorobenzenesulfonylamino)]butylazulene-1-sulfonic acid sodium salt (KT2-962), a potent and



long-acting NP-TRA.<sup>4</sup> As a continuation of our study in the structural requirement of KT2-962, an attempt was made to replace sulfonamide moiety to gain further information on the activity of NP-TRAs. We designed and synthesized various derivatives of KT2-962, replacing sulfonamide moiety with double amide and sulfonamide-amide, semicarbazone and thiosemicarbazone, inverse sulfonamide and N-sulfonylcarboxamide.

The target compounds were synthesized by methods A-D in Scheme 1. The azulene derivatives **1**, **4**, **7** and **11** were prepared from methyl 6-isopropyl-2-oxo-2H-cyclohept[b]furan-3-carboxylate according to the method reported by Takase.<sup>11</sup>

Double amides **2** and sulfonamide-amides **3** were synthesized by method A. Treatment of phthalimide **1** with hydrazine hydrate afforded the amine, which was then transformed into the double amides or sulfonamide-amides by condensation with the appropriate glycine derivatives in the presence of 1,3-dicyclohexylcarbodiimide. Hydrolysis of esters with 10% aqueous NaOH gave the carboxylic acids **2** and **3**.

Semicarbazones **5** and thiosemicarbazones **6** were synthesized by method B. Hydrolysis of the acetal **4** with 10% aqueous HCl followed by condensation with arylsemicarbazides or arylthiosemicarbazides in AcOH-H<sub>2</sub>O-THF, resulted in the formation of semicarbazones or thiosemicarbazones. Hydrolysis of esters gave the carboxylic acid sodium salts **5** and **6**.

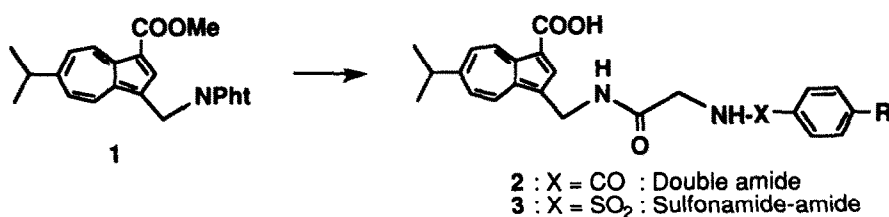
Inverse sulfonamides **9** and **10** were synthesized by method C. Deprotection of the tetrahydropyranyl ether of **7** with *p*-toluenesulfonic acid in MeOH followed by bromination of alcohols with PPh<sub>3</sub> and CBr<sub>4</sub> afforded the bromides **8**. Sulfonation of **8** with NaHSO<sub>3</sub> in aqueous dioxane provided the sodium sulfonates. Chlorination of sodium sulfonates with thionyl chloride followed by condensation with anilines gave the inverse sulfonamides. Hydrolysis of esters gave the carboxylic acids **9**. Sodium sulfonates **10** were obtained according to the reported method.<sup>12</sup>

N-sulfonylcarboxamides **13** and **16** were synthesized by method C. Hydrolysis of diester **11** with 10% aqueous NaOH at room temperature gave selectively the mono acid **12**. Condensation of **12** with arylsulfonamides in the presence of 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride and 4-di-

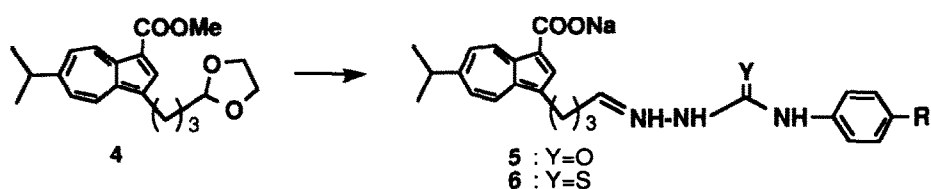
methylaminopyridine followed by hydrolysis of esters yielded the carboxylic acids **13**. Additionally, diester **11** was converted to the sodium sulfonates **16**. Demethoxycarbonylation of diester **11** by treatment with anhydrous phosphoric acid yielded **14**, which was then transformed into the N-sulfonylcarboxamides **15** by hydrolysis and condensation with arylsulfonamides. Sulfonation of **15** with pyridine-sulfur trioxide complex followed by treatment with sodium methoxide gave the sodium sulfonates **16**.

### Scheme 1. Modification of Sulfonamide Moiety in KT2-962

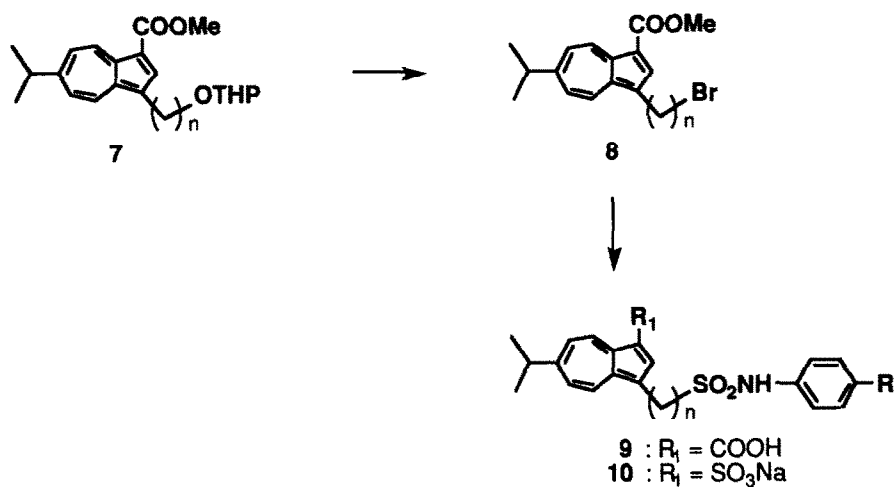
#### Method A : Double Amide and Sulfonamide - Amide Derivatives



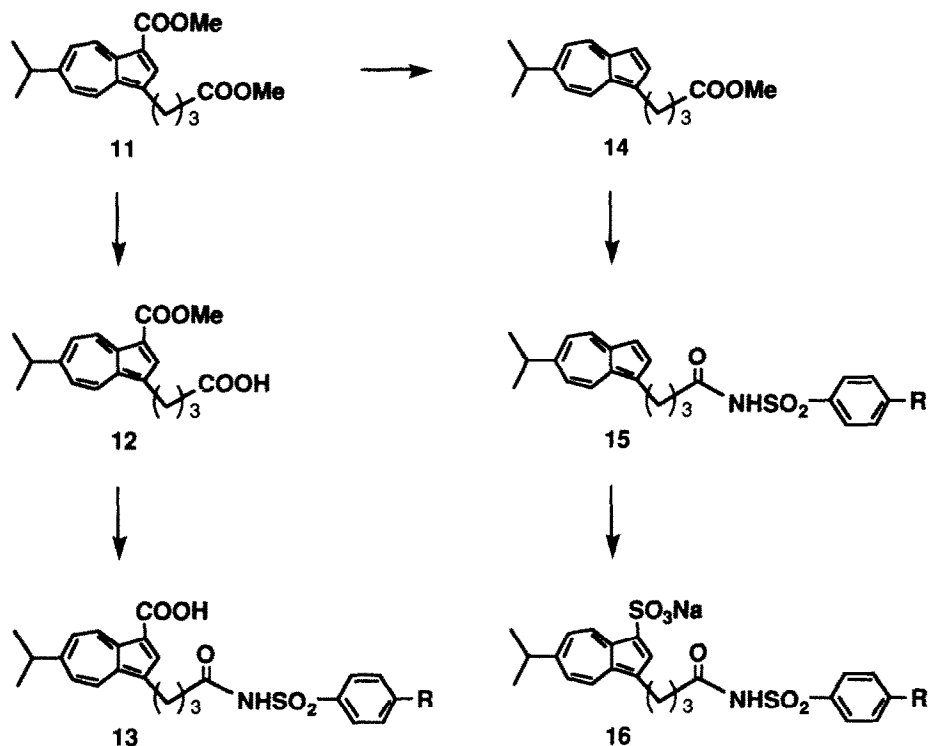
#### Method B : Semicarbazone and Thiosemicarbazone Derivatives



#### Method C : Inverse Sulfonamide Derivatives



## Method D : N-Sulfonylcarboxamide Derivatives



The synthetic compounds listed in Table 1 were compared in  $\text{TXA}_2$  antagonistic activities with KT2-962 and BM13,177 as reference compounds. They were tested for the effects on isolated rat thoracic aorta ( $\tau$ -receptor) precontracted by U-46619 ( $3.0 \times 10^{-8}$  M).<sup>13</sup> The concentrations that caused 50% relaxation are shown in Table 1. All tested compounds were found to be less potent than BM13,177, except for inverse sulfonamides **9f** and **10b-f**, with **10e** being the most potent, about 10 times more potent than BM13,177. Among inverse sulfonamide derivatives (compounds **9a-f** and **10a-f**), introduction of a chlorine atom into the para position of the phenyl ring (compounds **9d-f** and **10d-f**) increased the activity more than non-halogenated compound (compounds **9a-c** and **10a-c**). Replacement of the carboxyl group in the 1-position of the azulene ring with the sulfonic acid sodium salt (compounds **10a-f**) resulted in increased activity. The most active compound **10e** (the  $\text{IC}_{50}$  value of  $2.2 \times 10^{-7}$  M), however, had 3 orders of magnitude less activity as compared to KT2-962 (the  $\text{IC}_{50}$  value of  $9.0 \times 10^{-10}$  M). We also investigated the inhibitory effects of compounds **2a**, **3a**, **5a**, **6a**, **9e**, **10e**, **13b** and **16b** on platelet-rich plasma ( $\alpha$ -receptor) of rabbit.<sup>14</sup> The concentrations which cause 50% inhibition of the maximal aggregation are expressed as  $\text{IC}_{50}$  values and they are shown in Table 1. All compounds were less antagonistic for  $\alpha$ -receptors than BM13,177 and KT2-962 but **10e** was more selective antagonistic for  $\tau$ -receptors than BM13,177.

Squibb groups reported that the introduction of double amide<sup>9</sup> and semicarbazone<sup>10</sup> to P-TRAs, showed improvement in the inhibitory activity as compared to the allylic compound. However, these modifications

may not be applicable to NP-TRA such as KT2-962. The present study, therefore, indicates the importance of sulfonamide moiety for the activity of NP-TRA, KT2-962. Furthermore, non-peptide oxytocin<sup>15</sup> and fibrinogen<sup>16</sup> receptor antagonists, having sulfonamide moiety, were recently reported to increase non-covalent interactions at receptor sites. The study also suggests that there is a specific structural or non-covalent requirement for NP-TRAs which is different from that for P-TRAs.

**Table 1. Structures and TXA<sub>2</sub> Receptor Antagonistic Activities In Vitro of Azulene Derivatives**

comp.	R	n	mp °C	IC <sub>50</sub> (M) <sup>a</sup>	
				contraction <sup>b</sup>	aggregation <sup>c</sup>
<b>2a</b>	H	-	222 - 224	> 10 <sup>-5</sup>	> 10 <sup>-4</sup>
<b>2b</b>	Cl	-	223 - 225	> 10 <sup>-5</sup>	
<b>3a</b>	H	-	205 - 206	> 10 <sup>-5</sup>	> 10 <sup>-4</sup>
<b>3b</b>	Cl	-	195 - 197	> 10 <sup>-5</sup>	
<b>5a</b>	H	-	188 - 191	> 10 <sup>-5</sup>	> 10 <sup>-4</sup>
<b>5b</b>	Cl	-	169 - 171	> 10 <sup>-5</sup>	
<b>6a</b>	H	-	182 - 184	5.9 ± 0.1 × 10 <sup>-6</sup>	> 10 <sup>-4</sup>
<b>6b</b>	Cl	-	228 - 231	> 10 <sup>-5</sup>	
<b>9a</b>	H	3	145 - 146	1.2 ± 0.7 × 10 <sup>-5</sup>	
<b>9b</b>	H	4	177 - 178	8.7 ± 2.7 × 10 <sup>-6</sup>	
<b>9c</b>	H	5	189 - 190	1.5 ± 0.5 × 10 <sup>-6</sup>	
<b>9d</b>	Cl	3	143 - 144	9.1 ± 3.4 × 10 <sup>-6</sup>	
<b>9e</b>	Cl	4	175 - 176	2.6 ± 1.1 × 10 <sup>-6</sup>	> 10 <sup>-4</sup>
<b>9f</b>	Cl	5	171 - 172	1.7 ± 0.2 × 10 <sup>-6</sup>	
<b>10a</b>	H	3	171 - 172	8.7 ± 3.2 × 10 <sup>-6</sup>	
<b>10b</b>	H	4	216 - 218	1.5 ± 0.2 × 10 <sup>-6</sup>	
<b>10c</b>	H	5	212 - 213	1.4 ± 0.3 × 10 <sup>-6</sup>	
<b>10d</b>	Cl	3	222 - 223	8.6 ± 0.2 × 10 <sup>-7</sup>	
<b>10e</b>	Cl	4	206 - 207	2.2 ± 1.1 × 10 <sup>-7</sup>	> 10 <sup>-4</sup>
<b>10f</b>	Cl	5	207 - 208	8.6 ± 4.0 × 10 <sup>-7</sup>	
<b>13a</b>	H	-	173 - 175	> 10 <sup>-5</sup>	
<b>13b</b>	Cl	-	189 - 191	1.9 ± 0.2 × 10 <sup>-5</sup>	> 10 <sup>-4</sup>
<b>16a</b>	H	-	131 - 132	6.5 ± 1.3 × 10 <sup>-5</sup>	
<b>16b</b>	Cl	-	195 - 197	5.4 ± 0.1 × 10 <sup>-5</sup>	5.6 × 10 <sup>-5</sup>
<b>KT2-962</b>				9.0 ± 0.7 × 10 <sup>-10</sup>	8.7 × 10 <sup>-6</sup>
<b>BM13,177</b>				1.5 ± 0.1 × 10 <sup>-6</sup>	7.1 × 10 <sup>-6</sup>

<sup>a</sup> IC<sub>50</sub> values represent the mean ± SEM and calculated by regression analysis from the three dose groups of four different preparations. <sup>b</sup> Contraction of rat aorta was induced by 3.0 × 10<sup>-8</sup> M of U-46619. <sup>c</sup> Aggregation of rabbit platelet-rich plasma was induced by 4.0 × 10<sup>-6</sup> M of U-46619.

**Acknowledgment.** The authors would like to thank Prof. M. Shibasaki, University of Tokyo for encouragement throughout the present study.

**References and Notes**

1. Lefer, A. M. *DN and P.* **1989**, *25*, 265.
2. Lefer, A. M. *Drugs Today.* **1985**, *21*, 283.
3. (a) Hall, S.; Han, W-C.; Harvis, D. N.; Hedberg, A.; Ogletree, M. L. *J. Med. Chem.* **1989**, *32*, 974. (b) Hanasaki, K.; Arita, H. *Thromb. Res.* **1988**, *50*, 365. (c) Uski, T. K. *Acta. Physiol. Scand.* **1988**, *133*, 519. (d) Mckenniff, M.; Rodger, I. W.; Norman, P.; Gardiner, P. J. *Eur. J. Pharmacol.* **1988**, *153*, 149.
4. (a) Tomiyama, T.; Wakabayashi, S.; Kosakai, K.; Yokota, M. *J. Med. Chem.* **1990**, *33*, 2323. (b) Perzborn, E.; Seuter, F.; Fiedler, V. B.; Rosentreter, U.; Boshagen, H. *Arzneim.-Forsch./Drug Res.* **1989**, *39*, 1522. (c) Karasawa, A.; Shirakura, S.; Higo, K.; Kudo, K. *Arzneim.-Forsch./Drug Res.* **1991**, *41*, 1237.
5. Naka, M.; Hamanaka, M.; Sugioka, M.; Iwamura, H.; Sakata, M.; Kira, H.; Kegawa, T.; Kawasaki, A. *Biosignalling in Cardiac and Vascular System.* pp 422.
6. (a) Narisada, M.; Ohtani, M.; Watanabe, F.; Uchida, K.; Arita, H.; Doteuchi, M.; Hanasaki, K.; Kakushi, H.; Otani, K.; Hara, S. *J. Med. Chem.* **1988**, *31*, 1847. (b) Ohtani, M.; Narisada, M. *J. Med. Chem.* **1990**, *33*, 1027.
7. Bush, L. R.; Smith, S. G. *Thromb. Res.* **1988**, *44*, 377.
8. Resentreter, U.; Boshagen, H.; Seuter, F.; Perzborn, E.; Fiedler, V. B. *Arzneim.-Forsch./Drug Res.* **1989**, *39*, 1519.
9. Nakane, M.; Reid, J. A.; Han, W-C.; Das, J.; Truc, V. C.; Haslangen, M. F.; Garber, D.; Haris, D. N.; Hedberg, A.; Ogletree, M. L.; Hall, M. L. *J. Med. Chem.* **1990**, *33*, 2465.
10. Misra, R. N.; Brown, B. R.; Han, W-C.; Hedberg, A.; Webb, M. L.; Hall, S. E. *J. Med. Chem.* **1991**, *34*, 2882.
11. Yang, P. M.; Yasunami, M.; Takase, K. *Tetrahedron Lett.* **1971**, 4275.
12. Yanagisawa, T.; Wakabayashi, S.; Tomiyama, T.; Yasunami, M.; Takase, K. *Chem. Pharm. Bull.* **1988**, *36*, 641.
13. Thoracic aortas were obtained from male Wistar rats, and ring preparations were placed in organ bath containing Krebs solution (37°C, gassed with 5% CO<sub>2</sub> in O<sub>2</sub>). Each compound was added cumutively to the tissue contracted with U-46619 (3.0 × 10<sup>-8</sup> M), and then 50% relaxing concentration (IC<sub>50</sub>) for each compound was calculated.
14. Blood was obtained from male NZW rabbits and platelet-rich plasma (PRP, ab. 3 × 10<sup>8</sup> cell/ml) was prepared by centrifugation. Platelet aggregation was measured by the method of Born. The inhibitory effect of each compound on the platelet aggregation induced by U-46619 (4.0 × 10<sup>-6</sup> M) was examined and 50% inhibitory concentration (IC<sub>50</sub>) for each compound was calculate. see Born, G. V. R. *Nature (London)* **1962**, *194*, 927.
15. Evans, B. E.; Leighton, J. L.; Rittle, K. E.; Gilbert, K. F.; Lundell, G. F.; Gould, N. P.; Hobbs, D. W.; Dipardo, R. M.; Veber, D. F.; Pettibone, D. J.; Clineschmidt, B. V.; Anderson, P. S.; Freidinger, R. M. *J. Med. Chem.* **1992**, *35*, 3919.
16. Hartman, G. D.; Egbertson, M. S.; Halcenko, W.; Laswell, W. L.; Duggan, M. K.; Smith, R. L.; Naylor, A. M.; Manno, P. D.; Lynch, R. J.; Zhang, G.; Chang, C. T-C.; Gould, R. J. *J. Med. Chem.* **1992**, *35*, 4640.